



ZYMO RESEARCH



## Quick-RNA™ Viral 96 Kit

Viral RNA from any biological sample

### Highlights

- Spin-plate (96-well) purification of viral RNA from plasma, serum, urine, cell culture media, blood, saliva, cellular suspensions, swab, fecal and biopsy samples
- High-quality RNA is ready for Next-Gen sequencing, RT-qPCR, hybridization, etc.
- DNA/RNA Shield is included for sample collection, inactivation, storage and preservation.

Catalog Numbers:  
R1040, R1041



Scan with your smart-phone camera to  
view the online protocol/video.



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# Product Contents

<b>Quick-RNA™ Viral 96 Kit</b>	<b>R1040</b> (2 x 96 prep)	<b>R1041</b> (4 x 96 prep)
DNA/RNA Shield™ (2X concentrate)	125 ml	125 ml (x2)
Viral RNA Buffer <sup>1</sup>	100 ml (x2)	100 ml (x4)
Viral Wash Buffer <sup>2</sup> (concentrate)	48 ml	48 ml (x2)
DNase/RNase-Free Water	4 ml	4 ml (x2)
Zymo-Spin™ I-96 Plate	2	4
Collection Plate	2	4
Elution Plate	2	4
96-Well Plate Cover Foil	2	4
Instruction Manual	1 pc	1 pc

**Storage Temperature** - Store all kit components (i.e., buffers, columns) at room temperature.

Before use:

1 Add beta-mercaptoethanol (β-Me; user provided) to 0.5% (v/v) *i.e.*, add 500 µl β-Me per 100 ml **Viral RNA Buffer**.

2 Add 192 ml of 100% ethanol (204 ml of 95% ethanol) to the 48 ml **Viral Wash Buffer** concentrate.

# Specifications

- **Sample Sources** –  $\leq 400 \mu\text{l}$  plasma, serum, saliva, swab, urine, cell culture media, blood, cellular suspension, fecal sample or  $\leq 5 \text{ mg}$  biopsy sample.

For samples in UTM<sup>®</sup>/VTM<sup>®</sup>, PBS or saline, see Sample Preparation, page 5.

- **Purity** – RNA is ready for Next-Gen Sequencing, RT-qPCR, etc.
- **Binding Capacity** –  $10 \mu\text{g}$  total RNA (**Zymo-Spin<sup>™</sup> I-96 Plate**).
- **Elution Volume** –  $\geq 10 \mu\text{l}$  **DNase/RNase-Free Water**.
- **Equipment Needed** (user provided) – Beta-mercaptoethanol (b-Me), Ethanol (95-100%), Centrifuge with 96-well plate carrier.
- **Materials** (available separately) –

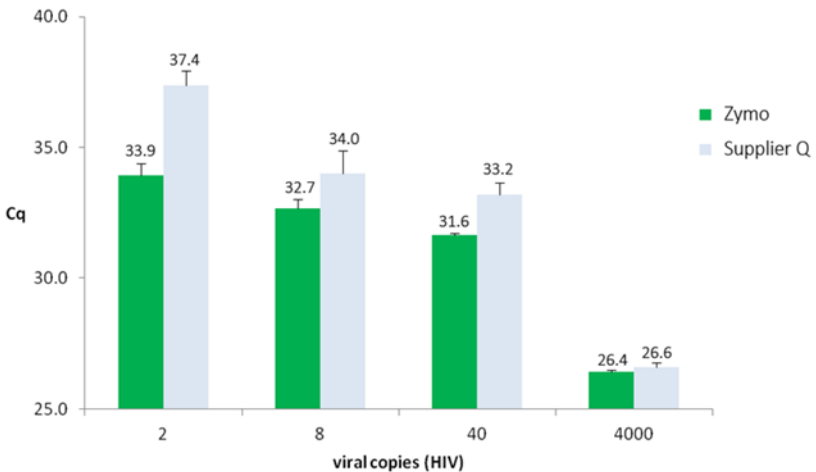
DNase I Set (E1010; 50 rxns.; 250 U DNase I (lyophilized) supplied w/ DNA Digestion Buffer, 4 ml)  
RNA Prep Buffer (R1060-2-50; 50 ml)  
RNA Wash Buffer (concentrate) (R1003-3-6, 6 ml)  
Proteinase K Set (D3001-2-20; 20 mg Proteinase K (lyophilized) supplied w/ Storage Buffer).

# Product Description

The **Quick-RNA™ Viral 96 Kit** is a spin-plate (96-well) purification of viral RNA from plasma, serum, urine, cell culture media, blood, saliva, cellular suspensions, biopsies, swab and fecal samples stored in **DNA/RNA Shield™** (for sample collection, nucleic acid preservation and inactivation of pathogens).

The kit also features a buffer system that facilitates complete viral particle lysis for efficient nucleic acid isolation. Small (> 50 nt) and large (> 200 kb) DNA and RNA are bound to each well of the plate, washed and eluted.

The isolated high-quality, total RNA is ready for all downstream applications such as Next-Gen sequencing, hybridization-based and RT-qPCR detection.



The **Quick-RNA™ Viral Kit** from Zymo Research ensures high sensitivity viral detection compared to that of Supplier Q. Viral RNA was isolated from plasma samples. Data shows the mean (+/- SD) of triplicate RT-qPCR measurements.

# Protocol

The protocol consists of: (I) Buffer Preparation, (II) Sample Preparation and (III) DNA/RNA Purification.

## (I) Buffer Preparation

- ✓ Add beta-mercaptoethanol (user provided) to 0.5% (v/v) i.e., add 500  $\mu$ l  $\beta$ -Me per 100 ml **Viral RNA Buffer**.
- ✓ Add 192 ml of 100% ethanol (204 ml of 95% ethanol) to the 48 ml **Viral Wash Buffer** concentrate.

## (II) Sample Preparation

- ✓ Perform all steps at room temperature (20-30°C).
- ✓ Up to 400 µl sample can be processed per prep.

**Samples in DNA/RNA Shield™<sup>1</sup> collection devices** (swabs, saliva, etc.)  
Proceed directly with purification, page 6.

**Swabs** (UTM®/VTM®, PBS, saline, etc.)

Proceed directly with purification, page 6.

Optional - To inactivate, store and preserve samples at room temperature prior to further processing, add **DNA/RNA Shield™**. See **Liquids**, below.

**Liquids** (plasma<sup>2</sup>, serum<sup>2</sup>, CSF, blood, saliva, urine, cell suspension, cell culture media)  
Add an equal volume of **DNA/RNA Shield™** (2X concentrate) to a volume of liquid sample (1:1) and mix well. Proceed with purification, page 6.

**Tissue**<sup>2</sup> (LCM, needle biopsy)

Add 400 µl **DNA/RNA Shield™** (1X) to a tissue sample (up to 5 mg) and mix well. Proceed with purification, page 6.

Optional - **Proteinase K treatment**<sup>3</sup> (protein-rich samples e.g., plasma, serum, saliva, sputum, tissue, can be treated). Materials sold separately.

Add 1% **Proteinase K** (v/v) at 20 mg/ml directly to a liquid sample. Mix well and incubate at room temperature for 15 minutes. Note: Up to 5% Proteinase K can be added (e.g., tissue). For example: Add 4-20 µl Proteinase K to each 400 µl sample.

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1 At this point, samples in DNA/RNA Shield™ can be stored at ambient temperature (4-25°C) for a month, 3 days at 37°C, or long-term (> 1 year) -20°C or below.

2 To remove particulate debris or cryoprecipitates (if any), centrifuge and transfer up to 400 µl of the cleared supernatant into a nuclease-free plate/tube (not provided).

3 Prior to use, reconstitute the lyophilized Proteinase K (D3001-2-20) and add 1,040 µl Storage Buffer. Mix well and store frozen aliquots.

### (III) RNA Purification

- ✓ Perform all steps at room temperature and centrifugation at 3,000-5,000 x g for 5 minutes.
- ✓ The sample input can be scaled up or down, proportionally.
- ✓ Do not use the **96-Well Cover Foil** on the spin-plate during RNA Purification. If necessary, use an Air Permeable Sealing Cover (#C2011-8); sold separately.

1. Add 800 µl **Viral RNA Buffer** to each 400 µl sample<sup>1</sup> (2:1) and mix well.
2. Transfer the mixture into each well of the **Zymo-Spin™ I-96 Plate**<sup>2</sup> mounted on a **Collection Plate** and centrifuge. Discard the flow-through from the collection plate.

Optional: At this point, DNase I treatment can be performed (see Appendices, page 7).

3. Add 500 µl **Viral Wash Buffer** to each well, centrifuge and discard the flow-through. Repeat this step.
4. Add 500 µl ethanol (95-100%) to each well and centrifuge. Then mount the spin-plate onto an **Elution Plate**.
5. To elute RNA, add 15 µl **DNase/RNase-Free Water** directly to the matrix of each well and centrifuge.

Alternatively, for highly concentrated RNA use ≥ 10 µl elution.

The eluted RNA<sup>3</sup> can be used immediately or stored frozen.

Use the **96-Well Cover Foil** to prevent the eluate from evaporation.

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1 Up to 400 µl sample can be processed per prep.

2 To process > 700 µl, the plate can be reloaded.

3 It is recommended to titrate the DNA/RNA eluate for downstream applications (i.e., RT/qPCR, etc.).



# Appendices

## DNase I Treatment

- ✓ For DNA-free RNA, DNase I treatment can be performed using DNase I Set (E1010; 50 reactions), RNA Prep Buffer (R1060-2-50) and RNA Wash Buffer (concentrate) (R1003-3-6); materials sold separately.

For each sample to be treated, prepare **DNase I Reaction Mix** in an RNase-free tube (not provided) and mix by gentle inversion:

<b>DNase I Reaction Mix</b>	
<b>DNA Digestion Buffer</b>	35 $\mu$ l
<b>DNase I</b> (reconstituted; 1 U/ $\mu$ l) <sup>1,2</sup>	5 $\mu$ l

1. Following RNA binding (page 6, step 2), add 400  $\mu$ l **RNA Wash Buffer**<sup>3</sup> to each well, centrifuge the plate and discard the flow-through.
2. Add 40  $\mu$ l **DNase I Reaction Mix** directly to the matrix of each well.
3. Incubate at room temperature for (20-30°C) for 15 minutes.
4. Add 500  $\mu$ l **RNA Prep Buffer** to each well, centrifuge the plate and discard the flow-through.
5. Proceed with RNA Purification (page 6, step 3).

1 Prior to use, reconstitute lyophilized 250 U **DNase I** (E1009-A) to 1U/ $\mu$ l (final concentration) with 275  $\mu$ l nuclease-free water (not provided), mix by gentle inversion and store frozen aliquots.

2 Unit definition – one unit increases the absorbance of a high molecular weight DNA solution at a rate of 0.001 A260 units/ml of reaction mixture at 25°C.

3 Before use, add 24 ml of 100% ethanol (26 ml of 95% ethanol) to the 6 ml **RNA Wash Buffer** concentrate.

# Ordering Information

Product Description	Catalog No.	Size
<b>Quick-RNA™ Viral 96 Kit</b>	R1040	2 x 96 preps.
	R1041	4 x 96 preps.

Individual Kit Components	Catalog No.	Amount
<b>DNA/RNA Shield™ (2X concentrate)</b>	R1200-25	25 ml
	R1200-125	125 ml
<b>Viral RNA Buffer</b>	R1034-1-50	50 ml
	R1034-1-100	100 ml
<b>Viral Wash Buffer (concentrate)</b>	R1034-2-24	24 ml
	R1034-2-48	48 ml
<b>Zymo-Spin I-96 Plate</b>	C2004	2
<b>Collection Plate</b>	C2002	2
<b>Elution Plate</b>	C2003	2
<b>DNase/RNase-Free Water</b>	W1001-30	30 ml
	W1001-100	100 ml
<b>DNA/RNA Shield™ Fecal Collection Tube</b>	R1101	10
<b>DNA/RNA Shield™ Collection Tube</b>	R1102	50
<b>DNA/RNA Shield™ Lysis Tube (microbe)</b>	R1103	50
<b>DNA/RNA Shield™ Lysis Tube (microbe) w/ swab</b>	R1104	50
<b>DNA/RNA Shield™ Lysis Tube (tissue)</b>	R1105	50
<b>DNA/RNA Shield™ Collection Tube w/ Swab (1 ml fill)</b>	R1106	10
	R1107	50
<b>DNA/RNA Shield™ Collection Tube w/ Swab (2 ml fill)</b>	R1108	10
	R1109	50
<b>DNA/RNA Shield™ Saliva Collection Kit (2 ml fill)</b>	R1210	1
<b>DNase I Set (250 U DNase I (lyophilized) supplied with DNA Digestion Buffer, 4 ml)</b>	E1010	1
<b>RNA Prep Buffer</b>	R1060-2-25	25 ml
	R1060-2-50	50 ml
<b>RNA Wash Buffer</b>	R1003-3-6	6 ml
	R1003-3-24	24 ml
<b>Proteinase K Set supplied w/ Storage Buffer</b>	D3001-2-5	5 mg
	D3001-2-20	20 mg

# Complete Your Workflow

- ✓ For sample collection, inactivation of pathogens, storage and preservation of nucleic acids, use DNA/RNA Shield™ collection devices:

DNA/RNA Shield™ Collection Devices	
DNA/RNA Shield™ Collection Tube w/ Swab (1 ml fill or 2 ml fill) #R1107, R1109	For swab samples of nasal, throat, etc.
DNA/RNA Shield™ Saliva Collection Kit (2 ml fill) #R1210	For saliva, sputum, etc.
DNA/RNA Shield™ Collection Tube DNA/RNA Shield™ Lysis Tube (microbe) DNA/RNA Shield™ Lysis Tube (microbe) w/ swab DNA/RNA Shield™ Lysis Tube (tissue) #R1102-R1105	For microbes, tissue, etc. (2 ml lysis tubes used for bead beating homogenization)

- ✓ For RNA clean-up (purification) from the aqueous phase (e.g., TRIzol, TRI Reagent or similar) or from any enzymatic reaction (e.g., DNase I treated RNA):

RNA Clean & Concentrator	
Microprep #R1013, R1015	DNase I Set included (#R1013)
MagBeads #R1081, R1082	(#R1082)

# Troubleshooting Guide

<b>Problem</b>	<b>Possible Causes and Suggested Solutions</b>
<b>RNA degradation</b>	<p>To prevent RNA degradation:</p> <p>Immediately collect and lyse fresh sample into a stabilization reagent (i.e., DNA/RNA Shield™) to ensure nucleic acid stability. Homogenized samples in DNA/RNA Shield™ can be stored frozen for later processing.</p>
<b>Low nucleic acid content and/or low sensitivity in downstream application</b>	<p>Incomplete deproteinization due to high-protein content in the sample (blood, plasma/serum, tissue etc.):</p> <ul style="list-style-type: none"><li>- Increase the volume of DNA/RNA Shield™ to the sample.</li><li>- Perform Proteinase K treatment (see Sample Preparation, page 4).</li></ul> <p>Increase eluate input:</p> <ul style="list-style-type: none"><li>-Titrate the DNA/RNA eluate for downstream applications (i.e., RT/qPCR).</li></ul>
<b>DNA contamination</b>	<p>To remove DNA:</p> <ul style="list-style-type: none"><li>- Perform DNase I treatment during the purification (page 6) or perform DNase I treatment post-purification (#R1080), then clean-up the treated sample.</li></ul>

For technical assistance, please contact 1-888-882-9682 or email [tech@zymoresearch.com](mailto:tech@zymoresearch.com)







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Integrity of kit components is guaranteed for up to one year from date of purchase. Reagents are routinely tested on a lot-to-lot basis to ensure they provide the highest performance and reliability.

This product is for research use only and should only be used by trained professionals. It is not for use in diagnostic procedures. Some reagents included with this kit are irritants. Wear protective gloves and eye protection. Follow the safety guidelines and rules enacted by your research institution or facility.

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