

# **Performance Evaluation of CAR-T Cell Source Isolated and Activated T Cells Using the ADAM-CellT**

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## Introduction

- Chimeric antigen receptor (CAR) T cell one of the immune cell therapy products, which is used for disease therapy such as cancer by overexpressing specific gene on T cells.
- Recently, CAR-T cells are being touted as the next generation cell to increase treatment efficiency<sup>1</sup>.
- For that reason, the importance of monitoring of the intermediate products has come
- The Coefficients of variation of precision within each measuring series did not exceed 10% **(A, D)**.

Results

- The analytical measurement range of the assays were 5 x  $10^4 \sim 4 \times 10^6$  cell/mL with ordinary least squares regression fit of y = 1.0196x - 27468 (r<sup>2</sup>=0.9962) (B).
- In the method comparison studies with BD LeucocountTM Kit, the correlation
- coefficient (r) was 0.994, and the slopes/intercepts were 1.008 (95% CI= 0.9836 to

into the limelight<sup>2</sup>.

- However, GMP facilities lack proper equipment which validates intermediate products, isolated or activated T cell, during CAR-T cell manufacturing processes<sup>3</sup>.
- ADAM-CellT has been developed with the purpose of monitoring the quantity and viability of intermediate products in CAR-T cell manufacturing processes and also it is an optimized product for accurately and quickly counting isolated and activated T cells.

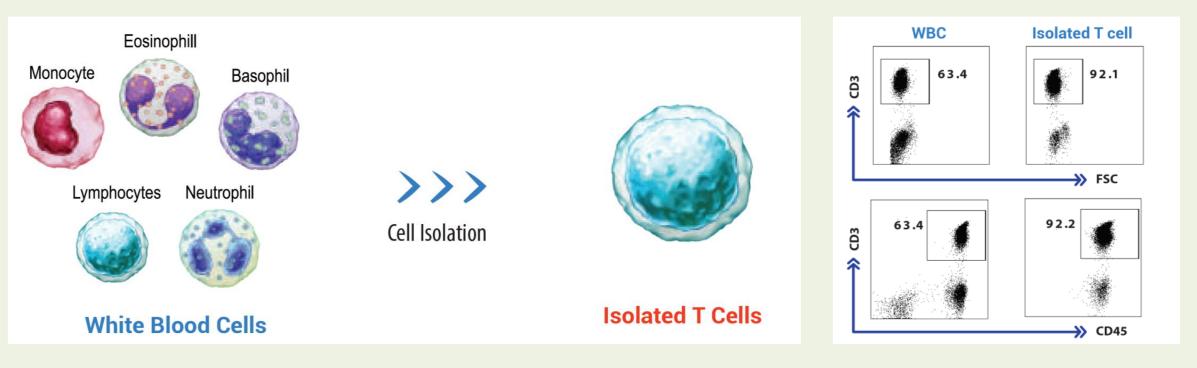
# **Methods and Procedures**

- The precision, linearity, and method comparison of ADAM-CellT were evaluated in accordance with CLSI guidelines EP05-A3, EP06-A, and EP09-A3.
- The isolated and activated T cells were made from human blood and were measured both with ADAM-CellT and by a comparative assay.
- The comparative assay was performed according to BD LeucocountTM Kit (BD) biosciences; 340523) protocol using flow cytometry.

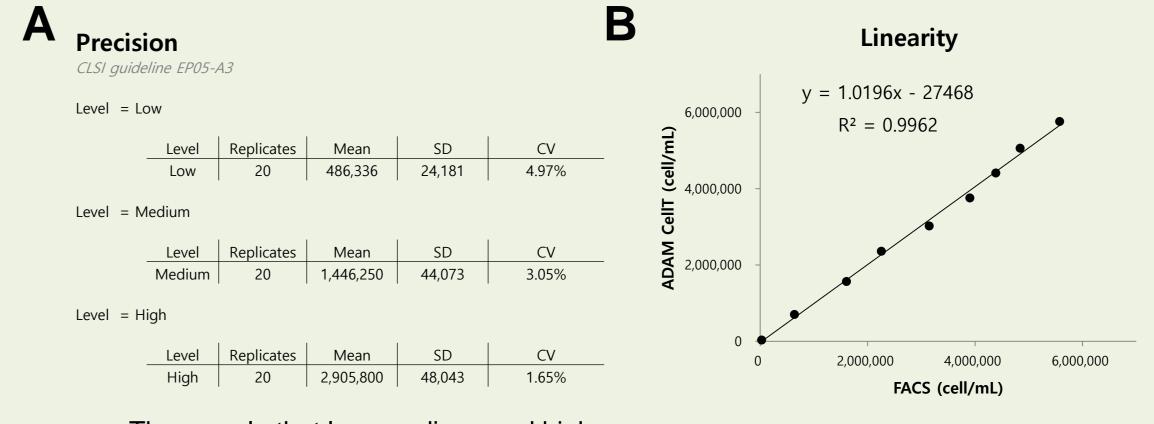
# **Optimization Workflow for Producing CAR-T**

1.029)/-114.2(95% CI= -45499 to 26091) by Passing-Bablock regression fit (C).

#### **Performance Evaluation in Isolated T Cells**



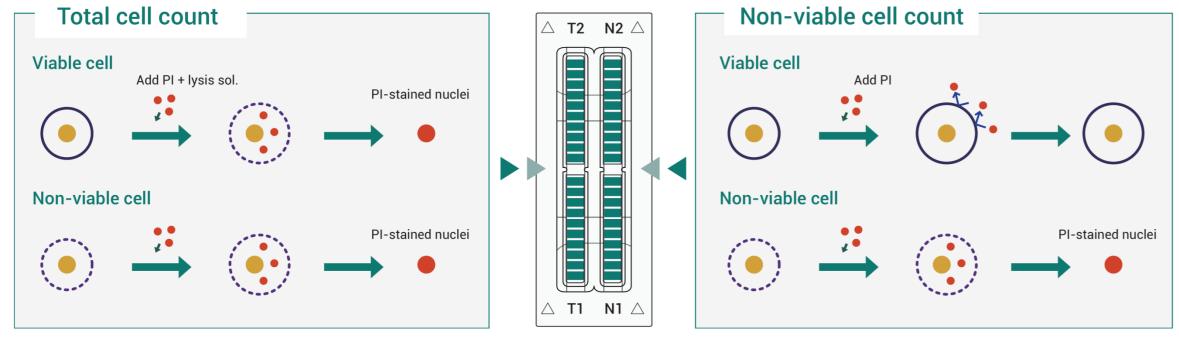
The profiles of T cells which was used for performance evaluation - Flow cytometric analysis of CD3 expression on unsorted (WBC; left panel) or sorted (Isolated T cell; right panel) human peripheral blood lymphocytes



The sample that low, medium, and high concentration of isolated T cells were counted with ADAM-CellT

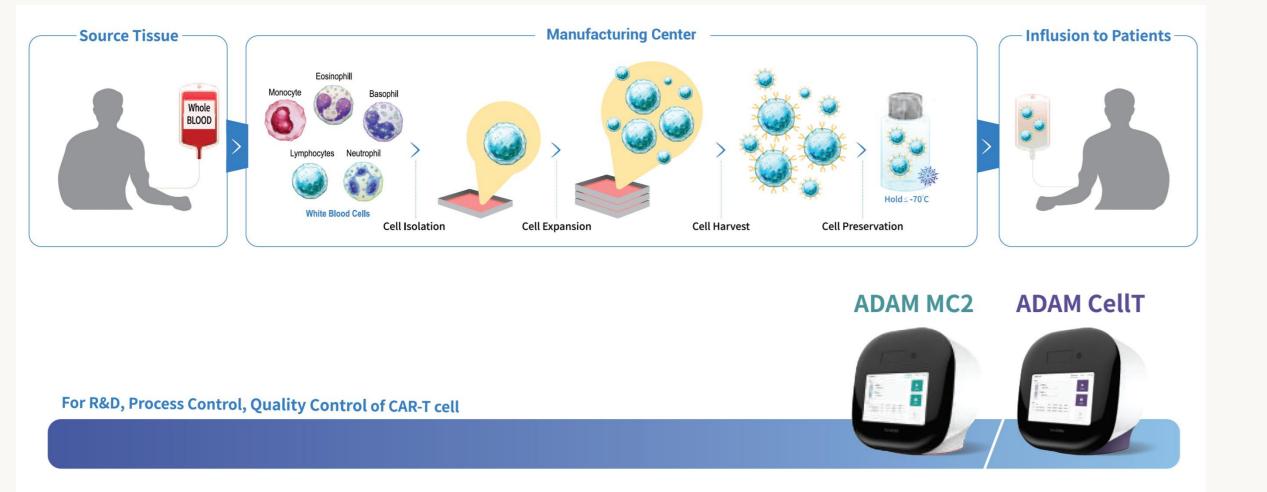
Comparison between flow cytometry and ADAM-CellT in Isolated T cells.

#### **Principle of Viability Measurement (PI-staining Method)**



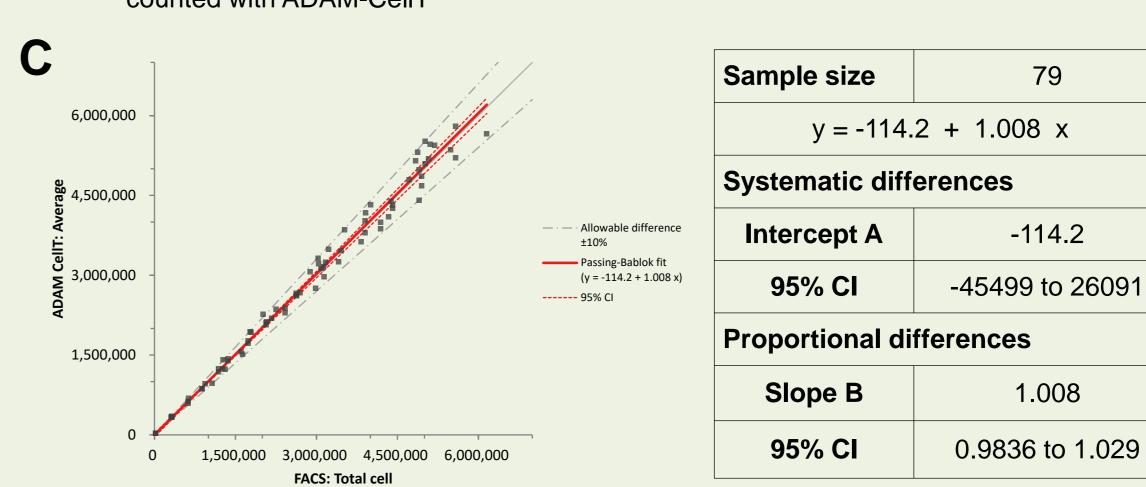
\* There are two types of disposable chips: 2 channel and 4 channel

#### **QC** Platform for Producing CAR-T



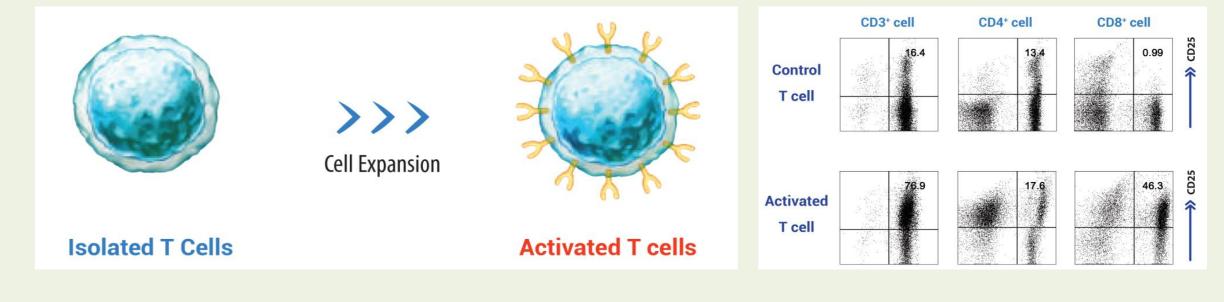
Monitoring the whole process from leukapheresis to the formulated product using the ADAM

• It is easy to monitor all different steps of the purification, expansion, and formulation



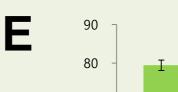
Correlation of T cell counting counting between flow cytometry and ADAM-CellT in Isolated T cells.

#### **Performance Evaluation in Activated T Cells**



The phenotypes of activated T cells which was used for performance evaluation - Flow cytometric analysis of CD25 expression on TCR stimulated (Activated T cell; bottom panel) or unstimulated (control T cell; upper panel) T cells.

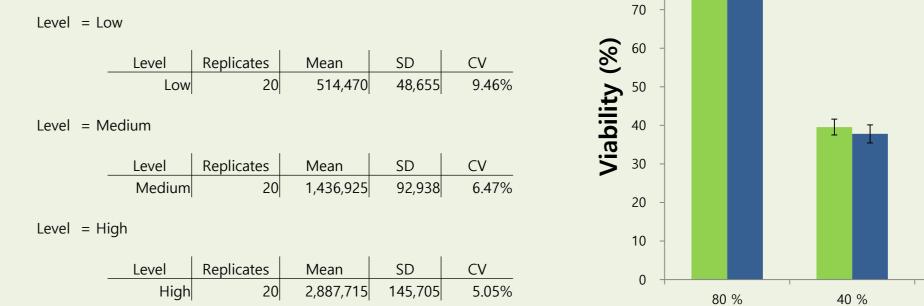
D Precision CLSI guideline EP05-A3

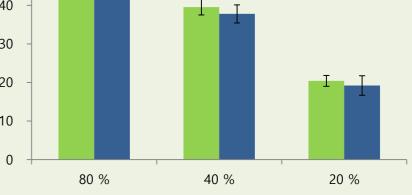


of CAR-T cells using the ADAM-MC2 and ADAM-CellT to ensure precise and reliable results. ADAM-MC2 and ADAM-CellT can be used for cGMP, process control and quality control of CAR-T cell manufacturing.



• Data indicates that the newly developed ADAM-CellT assay exhibits reliable performance. Consequently, we expect that ADAM-CellT will be a useful equipment to manage the monitoring of the quality of intermediate products during CAR-T cell manufacturing processes.





FACS

CellT

The sample that low, medium, and high concentration of activated T cells were counted with ADAM-CellT

Comparison of viability between flow cytometry and ADAM-CellT in Activated T cells

## References

1. Riddell SR. Cancer J. 20, 141-144 (2014). 2. Yonghong Li, *Engineering*. 5, 122-131 (2019). 3. Gee AP. Best Pract Res Clin Haematol. 31, 126-134 (2018).