Quick Start Protocol

GelRed® & GelGreen® Nucleic Acid Gel Stains

Catalog nos. 41002, 41003, 41004, 41005

Precast agarose gels

1. **Add GelRed® or GelGreen® to molten agarose at 1X final concentration.**
   For example, add 5 uL of 10,000X GelRed® or GelGreen® to 50 mL agarose.

2. **Cast gel and run samples.**
   Recommended loading is 50-200 ng DNA or ladder per lane, or 2-5 uL PCR product. If DNA concentration is unknown, run 1/2 to 1/3 the amount you would normally load on an EtBr gel. Overloading DNA can lead to poor band migration. If you need to load more DNA, use the post-staining protocol (other side).

3. **Image gel.**
   GelRed®: use UV light box with EtBr filter.
   GelGreen: use UV light box with SYBR® Green filter, or blue light illuminator such as a Dark Reader®.

Visit www.biotium.com for complete protocols, troubleshooting tips, and answers to frequently asked questions (FAQs).

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Post-staining agarose or acrylamide gels

1. **Run samples on gel with no DNA gel stain added.**

2. **Dilute GelRed® or GelGreen® in water at 3X final concentration.**
   For example, add 15 uL 10,000X GelRed® or GelGreen® to 50 mL water.

3. **Place gel in clean container with enough 3X gel stain to cover gel and incubate with rocking for 30 min.**
   Bands may begin to be detectable after 5 min.

4. **Image gel.**
   GelRed®: use UV light box with EtBr filter.
   GelGreen®: use UV light box with SYBR® Green filter, or blue light illuminator such as a Dark Reader®.

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